

Interaction between SARS-CoV-2 and human organism

¹Gheorghe Placinta, ¹Victor Pantea, ¹Lilia Cojuhari, ¹Valentin Cebotarescu, ²Lidia Placinta, ²Dan Croitoru

¹Department of Infectious Diseases, ²Department of Infectious Diseases, Tropical and Medical Parasitology
Nicolae Testemitanu State University of Medicine and Pharmacy, Chisinau, the Republic of Moldova

Authors' ORCID iDs, academic degrees and contribution are available at the end of the article

*Corresponding author: gheorghe.placinta@usmf.md

Manuscript received April 15, 2020; revised manuscript May 28, 2020; published online June 10, 2020

Abstract

Background: COVID-19 is a part of the betacoronaviridae group, sabercoviridae subgroup. At the moment we are confronting a pandemic, which has a completely new pathologic pattern for the whole world. Considering the highly contagious characteristics of the virus, which is the cause of death for many people, and also the fact that many people continue to be infected with this virus there is a stringing necessity of combating this pandemic. Because of that we need the knowledge that regards the physiopathologic mechanisms, particularities of the host-invader interactions, relevance of asymptomatic forms, explanations of the evolving divergencies, possibility of recurrent infections, clinical signs, comorbidities that harshen this pathology, pharmacologic agents that are effective to fight the infection and immune mechanisms of defense in the organism. A great importance is given for the identification of the initial criteria needed for a prognostic, to prevent the critical forms of pathology and to set the parameters for the severity indicators.

Conclusions: The research in the field of SARS-CoV-2 requires the gathering of the databases that are related to the investigated persons, to establish the clinico-evolutive differences for the COVID-19 patients according to many factors that could influence the course of the disease. An important difference is the identification of early signs and prevention of the critical disease forms, by extending the laboratory investigations, establishing the parameters for severity indicators through determining the degree of the immune response.

Key words: COVID-19, SARS-CoV-2, Coronavirus.

Cite this article

Placinta Gh, Pantea V, Cojuhari L, Cebotarescu V, Placinta L, Croitoru D. Interaction between SARS-CoV-2 and human organism. *Mold Med J.* 2020;63(2):57-62. doi: 10.5281/zenodo.3866031.

Introduction

Starting from 2020, the international medical community directed its attention towards the new coronavirus, COVID-19 and the pathology that it determines – SARS-CoV-2. This virus appeared for the first time in the city of Wuhan, Hubei province, China and has spread very fast at a global level. The hastened spread of this virus puts the world into the evidence of the high transmissibility of this family of viruses with a significant mortality and morbidity. As this virus appeared recently, it needs a continuous research. We are in a continuous pandemic with COVID-19, scientists are fighting to understand the similarities and differences between COVID-19 and SARS-CoV-1 at genomic and transcriptomic level [1]. The main targets of the virus are the lungs, but it has a serious effect on heart especially in case of a cronically affected one that needs a supplementary effort in order to deliver the oxygenated blood to the tissues. Patients with persistent cardiac affections have a weaker immune system. The immune response is decreased when advancing in age and in people with chronic comorbidities. One of the objectives of this study was to present the most recent data in order to determine the immune response in patients with COVID-19.

To realize this objective we used the Google Search results and the PubMed database. For advanced search of

the bibliographic sources we applied the following filters: articles with integral texts, articles in the English language published before 28.04.2020. After a preliminary analysis of the titles we selected the original, editorial articles of narrative synthesis, systematic and meta-analysis that contained relevant information and contemporary concepts about COVID-19. The information of the publications included in the bibliography was collected, classified, evaluated and synthesized putting into evidence the main aspects of the contemporary visions about the infection and immunity in COVID-19.

Results and discussion

The SARS-CoV-2 virus is entering the organism via the ACE2 [2-10] that is intensely expressed in the cells of the nasal mucosa, bronchi, lungs (type I and type II alveolocytes), heart, esophagus, kidneys, stomach, biliar sac, ileon, and neurons [5, 7, 8, 10]. After the invasions, the cellular proteases (TMPRSS2 and Furing) will lead to the final internalisation of the virion [11].

The anterior sites are the most affected in case of SARS-CoV-2, the epithelial cells being distructed after the invasion [12]. The supposed enter gates are – the respiratory epithelia at pulmonary level [7], through the olfactory filia that are penetrating the lamina cribrosa [5, 13]

or through the nervous fibers of the pulmonary mechano- and chemo-receptors via the nucleus *tracti solitarii* and the neuroinvasion can also take place through the lymphatics [13]. The specificity for this receptor was conferred by the spike glycoprotein that has the S1 and S2 subunits that are attached to the viral envelope (capsid) [1, 4, 5, 14, 15]. There are numerous polymorphisms for this protein in the same viral species, thing also characteristic for COVID-19 [1], the spike protein is expressed 10-20 times more intensely by COVID-19 comparatively with SARS-CoV-2, the fact argued by the enhanced viral permeability [5, 16]. Another way of cell invasion is through the CD147 receptor (Basigin/EMMPRIN) via the spike-CD147 protein [17], another membranary receptor that could stimulate the matrix metalloproteinases (MMP) – MMP1, MMP2, MMP3, MMP9 [18, 19], because of this COVID-19 could affect the T lymphocytes [20]. Once entered in the cell, the virus starts the translation of the own ARN for the pp1a and pp1ab proteins codified by the ORF1a and ORF1b genes – that play a key role in the viral replication, that are cleaved by the nsp 1-16 proteins [14, 21], respectively the inhibition of this chain could significantly decrease the infecting potential in the host. The antigens characteristic for COVID-19 can be recognized by the following receptors in human body – Toll-like receptor (TLR), Nod-like receptor (NLR), pattern recognition receptors (PRR), RIG-1 like receptor (RLR) and the cytosolic receptor for melanoma differentiation-associated gene 5 (MDA5) [4, 14] that are on the surface of the antigen presenting cells and also interact with the CD26, cyclophilin enzyme [15]. This leads to the release of the pathogen-associated molecular patterns (PAMP) [14], that could interact with the cells of the human organism in order to unleash the formation of inflammasomes that lead to the apoptosis through the NF- κ B pathway intracellularly and will determine the local inflammation of tissues that in the most severe cases will determine the systemic inflammatory response syndrome (SIRS) [22], along with the acute respiratory distress syndrome (ARDS), thrombosis and pulmonary embolism [12]. In the infection with SARS-CoV-2 we observe a hyperglycemia, referring to the infection with SARS-CoV-2 we haven't observed such modification at the moment [11]. During the febrile syndrome manifestation eczematous eruptions may appear [23, 24] or vesicles surrounded by erythematous halos that could form crusts – that are located preponderantly in the thoracic region [25], [42]. Lesions characteristic for frostbites were identified for the most patients with SARS-CoV-2 without confirming their association with the respective infection [26]. More recently we have a report of the acral cutaneous lesions [27].

An incidence of 1% of conjunctivitis was reported in a meta-analysis [28]. Along with the spike protein, COVID-19 presents the hemagglutinin-esterase, membranary protein M, nucleocapsidic protein, the small capsidic protein, the internal protein and specific group protein – potential targets for vaccines [29].

In SARS-CoV-2 the activation of the CD8+ lymphocytes and CD4+ lymphocytes is being observed along with the

C3a and C5a components of the complement that play a major role here, the worst scenario being the systemic inflammatory response syndrome (SIRS) with multiple organ dysfunction syndrome (MODS) [1, 2, 4]. In SARS-CoV-2 was observed the cytokine storm [10, 30]. In the plasma of the patients with SARS-CoV-2 we identified the following cytokines – IL-1, IL-2, IL-4, IL-6, IL-7, IL-10, IL-12, IL-13, IL-17 and the colony stimulating factor of granulocytes (G-CSF), the macrophage colony stimulating factor (M-CSF), IP-10, MCP-1, MIP-1 α , the hepatocyte growth factor (HGF), IFN- γ and TNF- α [13, 14], particularly we see the presence of the interferon 1 (IFN1) that plays a major role in the inhibition of the Th1 differentiation and the amplification of the Th2 cells, chemokines like IP-10 and MCP-1 that are intensely expressed in the COVID-19 [13, 30] will determine the chemotaxis of the numerous cells of the immune system, especially the neutrophils that will cause diffuse alveolar lesions and ARDS [1-3, 10, 13, 30]. The severe cases have a basal lymphopenia, leucocytosis an increased neutrophil/lymphocyte level and less monocytes, eosinophils and basophils. The number of Th lymphocytes is decreased, the number of T_s lymphocytes is decreased, the number of naive Th circulant levels is increased, the number of Th lymphocytes is decreased [9, 31] this is explained by the fact that COVID-19 could enter these cells via the CD147 [20]. Th lymphocytes express the Notch1 gene [32], having a high fraction of fatigued T lymphocytes [12]. The serine proteases and cathepsins are released during the immune response and could cleave the spike proteins – reducing the viral permeability [14]. The levels of Angiotensin II is positively correlated with the viral dose and the severity of lesions because it denotes the high levels of ACE receptors expression and hastens the ARDS installation [3]. The ORF1ab, ORF10 and ORF3a proteins could coordinate the hem attack in the β 1 chain of the hemoglobin and "steal" the Fe ions from the hem, also the capsid glycoproteins could bind to the porphyrinic chain [33], a meta-analysis has shown that patients with SARS-CoV-2 have a decreased total hemoglobin [34]. An increased level of C reactive protein (PCR) is observed because of the systemic inflammation, an increased level of ferritin because of the hem attack and an increased level of procalcitonin due to the response to the infection in the organism [35]. At the cardiac level an acute cardiac lesions could appear [36], that could evolve in a cardiogenic shock having an elevated concentration of troponin. The mechanisms of proposed lesions are – myocardial hypoxia, the destruction of the microcirculation vessels, endothelial desquamation and cytokine mediated lesions – there are no proofs at the moment that the myocardium could be infiltrated with lymphocytes, the global effects are represented by the cardiac failure, arrhythmia and cardiac decompensation [29]. Taking into consideration that the dopamin decarboxylase gene is positively correlated with the ACE2 gene, we could suppose that alterations in the synthesis of the dopamine are correlated with the physiopathologic mechanisms of the COVID-19 [37]. At the initial stages it could cause anosmia, ataxia and convulsions – in the most severe cases it could lead to respiratory stop [38].

The infection starts with the activation of the dendritic cells that reside in the pulmonary tissue, that will present further the antigens to the naive T cells that will secrete different chemokines and cytokines [29], the CD8+ cells have the tendency to secrete high quantities of IL-6 and the CD4+ have the tendency to secrete high quantities of IFN- γ and GM-CSF [12, 29] but they could secrete the chemokine ligand 9,10 and 11 (CXCL) and cytolytic molecules just as granzyme B [29]. A series of comorbidities could make the infection with SARS-CoV-2 worse, like arterial hypertension, determined by the amplification of the renin-angiotensin-aldosterone system (RAAS) by inducing endothelial lesions that will lead to cardiac, pulmonary and renal complications [3, 14, 39]. It can induce the destruction of the hemato-encephalic barrier; the cerebrovascular pathologies will worsen the SARS-CoV-2 flow [5, 39]. The pulmonary pathologies (Chronic obstructive pulmonary diseases – COPD), diabetes mellitus and advanced age were recognized as risk factors [7, 11, 14], in diabetes mellitus an increased expression of ACE2, furin was observed along with the dysfunction of the CD4+ cells and increased concentration of IL-6 [11], furin being codified by the Notch1 gene [32]. The patients with respiratory insufficiency need on-stage intubation [40]. The infection has weaker manifestations in children because they are temporary adapted in order to combat the viral infections, also the ACE2 receptors are less pronounced in their organism [9, 12], the male gender has a more severe symptomatic because of increased ACE2 expression [12]. Patients with pulmonary cancer are more susceptible to COVID-19 infection [41]. We have to mention that along with the generic symptoms of SARS-CoV, although we mentioned before about dermatological findings, we can observe chicken-pox like vesicles [42] and an increased level of lactate-dehydrogenase (LDH) and creatinofosokinase (CPK) [3]. We reported a case in which an asymptomatic woman that presented an infection with COVID-19 in the tardive partum period hasn't transmitted the virus vertically [43].

The long-term consequences after SARS-CoV-2 infection at the moment are not known in details, but it was reported that in 40% of the patients with SARS-COVID-1 were observed cardiovascular anomalies and dysregulations of the lipidic metabolism [44].

A series of pharmacological agents are potentially efficient for combating SARS-CoV-2 infection, we will expose a part of them: Baricitinib [30, 45], Ruxolitinib and Fedratinib are inhibitors of JAK-STAT signaling and respectively it wouldn't diminish the systemic inflammatory effects in SARS-CoV-2 [45]. Chloroquine is supposed to be a strong antiviral agent, useful in the treatment of pneumonia caused by COVID-19, being very economic from the financial point of view [9, 21, 30, 46, 47], it is very effective to inhibit the destruction of hem by the ORF1ab, ORF3a and ORF6 proteins along with the capsid glycoproteins [33]. Hidroxychloroquine (HCQ) could glycosilate the ACE2 receptors, it could induce the cleavage of the spike proteins and block the cytokine storm, but the Ivermectin – an

tiparasitary agent could interfere with the nuclear transport of the viral proteins dependent on the α/β importin, a combination of HCQ/Ivermectin is recognized to be used in order to combat SARS-CoV-2 but we haven't conducted a study about its effect [48]. The efficiency of Ivermectin in SARS-CoV-2 was tested only *in vitro* at the moment (14.04.2020) [49]. Erythropoietin has shown its efficiency in order to combat the enemy caused by SARS-CoV-2 in case study [50], also in a theoretic review there are proposed the pharmacological agents used in case of pulmonary edema caused by high altitudes (EPAM) – acetazolamide, nifedipine and inhibitors of phosphodiesterase without being clinically tested at the moment [51]. The inhibitors of the viral proteases (nsp inhibitors) like lopinavir and ritonavir are potential antiviral agents that could combat this infection [21], a combination of lopinavir-ritonavir was used to combat SARS-COVID-1 and MERS-COVID [52]. Favipiravir could inhibit the binding of the spike and ORF7a proteins to the porphyrin ring [33]. Corticosteroids are potential anti-inflammatory agents [30], but the corticosteroids determine an increase in the spread of the virus into the organism [13]. The inhibitors of the interleukin-6 could be useful in order to combat SARS-CoV-2 [9], the results of a clinical study show that the administration of tocilizumab (an inhibitor of IL-6) could be useful in order to combat COVID-19 if it is administered many times [53]. At the moment the inhibition of Notch1 gene is discussed in order to prevent the cytokine storm [32]. A study made *in vitro* proves that the multiplication of COVID-19 is inhibited via the blockage of CD147 with agents like Meplazumab [17, 54]. Azithromycin can inhibit the action of some matrix metalloproteinases. AS the STEM cells present a CD147 receptor, they could also serve as a target for SARS-CoV-2 [18], being proved to be a good adjuvant agent along with hydrochloroquine in a non-randomised clinical study [55].

The identification of the viral dose from the salivary prelevate taken from the posterior oropharyngeal mucosa could be an efficient method to determine COVID-19 at an incipient stage of manifesting SARS-CoV-2, but it is necessary to combine this inspection with an antibody analysis because they present an inverse correlation with the viral dose [52]. In the blood plasma consequently there are antibodies with a median seroconversion on the 11th day, IgM with a median seroconversion on the 12th day and IgG with a median seroconversion on the 14th day ($p < 0.05$), the tests for the antibodies could be useful in order to assess clinical suspicions in the symptomatic patients that don't have a viral ARN detectable in plasma [56]. The administration of convalescent plasma for the patients with SARS-CoV-2 was proven to be efficient to attenuate the infection manifestations, according to a theoretic review in a series of cases, at the moment there are no available randomised clinical trials (RCT) [57]. The antibodies in SARS-CoV-2 infection could be detected only in the medium and late stage [58]. The patients that could recover after an infection of SARS-CoV-2 could have high viral doses in the fecal masses [59].

At the moment the histopathological aspects of SARS-CoV-2 are not yet known in details, according to a theoretic review, the patients with pulmonary cancer presented histopathological patterns- macroscopic (pleurisy, pericarditis, pulmonary consolidations and edema) and microscopic (alveolar proteic exsudates, focal reactive hiperplasia of the pneumocytes with grouped cellular infiltrations and gigantic polynucleated cells without hyalin membranes). The patients that presented the ground glass image on the pulmonary radiography, had diffuse alveolar lesions with exsudates (characteristic for the distant lesions of immune systems in SIRS). No viral inclusions were detected in the histopathological findings [60]. Diffuse alveolar lesions were observed in another study, along with infiltrates of CD8+ lymphocytes and CD4+ lymphocytes around the high caliber bronchioli, in the majority of cases being present hemoragic foci and intracellular trombi, type II pneumocytes with cytopatic effect (desquamated), the alveolar capillaries were hypertrophiated with perivascular edema, in this case there was no hyaline membrane but were identified CD61+ megacariocytes. In the heart we identified disseminated cellular necrosis, without focal accumulations [61]. The ground glass image in the pulmonary radiography identified diffuse alveolar lesions in the organizatoric phase, fibrinous alveolar proteic exsudate, hyperplasy of the reactive pneumocytes type II along with interstitial fibrosis and chronic inflammatory infiltrations [62]. At the renal level there were acute lesions of the proximal tubule manifested with vacuolar degeneration and epithelium desquamation. For the patients with pathologic pulmonary manifestation we identified the acute pyelonephritis, more rarely – erythrocytic aggregation in the tubes with consequent obstruction, occasionally we observed the hemosiderin inclusions. The distal tubules didn't present serious pathological modifications [63].

Conclusions

SARS-CoV-2 became a global threat for health, with the number of patients increasing gradually. An increased incidence of diverse comorbidities was observed along with those that presented severe infection. The mechanism of cardiac lesions is not clear but it probably implies a combination of direct viral lesions and immunised damage mediated by the inflammatory cytokines/chemokines and cytotoxic response of the immune cells in the further infection stages. The immune response of the host and the contributors to the cytokine storm in the infection with SARS-CoV-2 is complex. The exhaustion and significant deregulation of the T lymphocytes could contribute to the immune degradation and hyperactivity. The treatments for COVID-19 are bimodal with a treatment group oriented towards the temporary infection and viral replication, there is also another group that takes into consideration the modulation of the immunity at the systemic inflammatory stage. It is important to make complete immune searches for the patients with diverse comorbidities in order to understand the systemic deregulations better. It is necessary to acquire

more data about the immune response according to the severity of disease and to take the needed measures in the managerial acts in the infection control.

References

1. Ciotti M, Angeletti S, Minieri M, et al. COVID-19 outbreak: an overview. *Chemotherapy*. 2020 Apr 7:1-9. doi: 10.1159/000507423.
2. Wu YC, Chen CS, Chan YJ. The outbreak of COVID-19: an overview. *J Chin Med Assoc*. 2020;83(3):217-220. doi: 10.1097/JCMA.000000000000270.
3. Guo J, Huang Z, Lin L, Lv J. Coronavirus disease 2019 (COVID-19) and cardiovascular disease: a viewpoint on the potential influence of angiotensin-converting enzyme inhibitors/angiotensin receptor blockers on onset and severity of Severe Acute Respiratory Syndrome Coronavirus 2 infection. *J Am Heart Assoc*. 2020;9(7):e016219. doi: 10.1161/JAHA.120.016219.
4. Yi Y, Lagnition PNP, Ye S, Li E, Xu RH. COVID-19: what has been learned and to be learned about the novel coronavirus disease. *Int J Biol Sci*. 2020;16(10):1753-1766. doi: 10.7150/ijbs.45134.
5. Baig AM, Khaleeq A, Ali U, Syeda H. Evidence of the COVID-19 virus targeting the CNS: tissue distribution, host-virus interaction, and proposed neurotropic mechanisms. *ACS Chem Neurosci*. 2020 Apr 1;11(7):995-998. doi: 10.1021/acscchemneuro.0c00122.
6. Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270-273. doi.org/10.1038/s41586-020-2012-7.
7. Weston S, Frieman MB. COVID-19: knowns, unknowns, and questions. *mSphere*. 2020 Mar 18;5(2). pii: e00203-20. doi: 10.1128/mSphere.00203-20.
8. Sun P, Lu X, Xu C, Sun W, Pan B. Understanding of COVID-19 based on current evidence. *J Med Virol*. 2020 Feb 25. doi:10.1002/jmv.25722. [Epub ahead of print].
9. Ludvigsson JW. Systematic review of COVID-19 in children shows milder cases and a better prognosis than in adults. *Acta Paediatr*. 2020 Mar 23. doi: 10.1111/apa.15270. [Epub ahead of print].
10. Jin Y, Yang H, Ji W, et al. Virology, epidemiology, pathogenesis, and control of COVID-19. *Viruses*. 2020;12(4):1-17. doi: 10.3390/v12040372.
11. Sing AK, Gupta R, Ghosh A, Misra A. Diabetes in COVID-19: prevalence, pathophysiology, prognosis and practical considerations. *Diabetes Metab Syndr Clin Res Rev*. 2020;14(4):303-310. doi: 10.1016/j.dsx.2020.04.004.
12. Yuki KS, Fujiogi M, Koutsogiannaki S. COVID-19 pathophysiology: a review. *Clin Immunol*. 2020;215:108427. doi: 10.1016/j.clim.2020.108427.
13. Li YC, Bai WZ, Hashikawa T. The neuroinvasive potential of SARS-CoV2 may be at least partially responsible for the respiratory failure of COVID-19 patients. *J Med Virol*. 2020 Feb 27. doi: 10.1002/jmv.25728. [Epub ahead of print].
14. Guo YR, Cao QD, Hong ZS, et al. The origin, transmission and clinical therapies of coronavirus disease 2019 (COVID-19) outbreak - an update on the status. *Mil Med Res*. 2020;7(1):11. doi: 10.1186/s40779-020-00240-0.
15. Vankadari N, Wilce JA. Emerging WuHan (COVID-19) coronavirus: glycan shield and structure prediction of spike glycoprotein and its interaction with human CD26. *Emerg Microbes Infect*. 2020;9(1):601-604. doi: 10.1080/22221751.2020.1739565.
16. Liu Y, Gayle AA, Wilder-Smith A, Rocklöv J. The reproductive number of COVID-19 is higher compared to SARS coronavirus. *J Travel Med*. 2020;27(2):1-4. doi: 10.1093/jtm/taaa021.
17. Wang K, Chen W, Zhou YS, et al. SARS-CoV-2 invades host cells via a novel route: CD147-spike protein. *BiorXiv*. 2020 Mar 14. Preprint doi: 10.1101/2020.03.14.988345.
18. Ulrich H, Pillat MM. CD147 as a target for COVID-19 treatment : suggested effects of azithromycin and stem cell engagement. *Stem Cell Rev Rep*. 2020 Apr 20. doi: 10.1007/s12015-020-09976-7. [Epub ahead of print].

19. Hasaneen NA, Cao J, Pulkoski-Gross A, Zucker S, Foda HD. Extracellular Matrix Metalloproteinase Inducer (EMMPRN) promotes lung fibroblast proliferation, survival and differentiation to myofibroblasts. *Respir Res.* 2016;17:1-14. doi: 10.1186/s12931-016-0334-7.
20. Wang X, Xu W, Hu G, et al. SARS-CoV-2 infects T lymphocytes through its spike protein-mediated membrane fusion. *Cell Mol Immunol.* 2020 Apr 7. doi: 10.1038/s41423-020-0424-9. [Epub ahead of print].
21. Ahn DG, Shin HJ, Kim MH, et al. Current status of epidemiology, diagnosis, therapeutics, and vaccines for novel coronavirus disease 2019 (COVID-19). *J Microbiol Biotechnol.* 2020;30(3):313-324. doi: 10.1021/acscentsci.0c00272.
22. Jo E, Kim JK, Shin D, Sasakawa C. Molecular mechanisms regulating NLRP3 inflammasome activation. *Cell Mol Immunol.* 2016;13(2):160-169. doi: 10.1038/cmi.2015.95.
23. Amatore F, Macagno N, Mailhe M, Demarez B, Gaudy-Marqueste C, Grob JJ, et al. SARS-CoV-2 infection presenting as a febrile rash. *J Eur Acad Dermatol Venereol.* 2020 Apr 24. doi: 10.1111/jdv.16528. [Epub ahead of print].
24. Avellana Moreno R, Villa E, Avellana Moreno V, Estela Villa C, Aparicio M. Cutaneous manifestation of COVID-19 in images: a case report. *J Eur Acad Dermatol Venereol.* 2020 Apr 24. doi: 10.1111/jdv.16531. [Epub ahead of print].
25. Tammaro A, Adebajo GAR, Parisella FR, Pezzuto A. Cutaneous manifestations in COVID-19: the experiences of Barcelona and Rome. *J Eur Acad Dermatol Venereol.* 2020 Apr 24. doi: 10.1111/jdv.16530. [Epub ahead of print].
26. Piccolo V, Neri I, Filippeschi C, et al. Chilblain-like lesions during COVID-19 epidemic: a preliminary study on 63 patients. 2020 Apr 24. doi: org/10.1111/jdv.16526. [Epub ahead of print].
27. Recalcati S, Barbagallo T, Frasin LA, Prestinari F, Cogliardi A, Provero MC, Dainese E, Vanzati A. Acral cutaneous lesions in the Time of COVID-19. *J Eur Acad Dermatol Venereol.* 2020 Apr 24. doi: 10.1080/09546634.2020.1764904. [Epub ahead of print].
28. Loffredo L, Pacella F, Pacella E, Tiscione G, Oliva A, Violi F. Conjunctivitis and COVID-19: a meta-analysis. *J Med Virol.* 2020 Apr 24. doi: 10.1002/jmv.25938. [Epub ahead of print].
29. Zhu H, Rhee JW, Cheng P, et al. Cardiovascular complications in patients with COVID-19: consequences of viral toxicities and host immune response. *Curr Cardiol Rep.* 2020;22(5):32. doi: 10.1007/s11886-020-01292-3.
30. Sarzi-Puttini P, Giorgi V, Sirotti S, et al. COVID-19, cytokines and immunosuppression: what can we learn from severe acute respiratory syndrome? *Clin Exp Rheumatol.* 2020;38(2):337-342.
31. Qin CP, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, Xie C, Ma K, Shang K, Wang W, Tian DS. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clin Infect Dis.* 2020 Mar 12. doi: 10.1093/cid/ciaa248. [Epub ahead of print].
32. Rizzo P, Vieceli Dalla Sega F, Fortini F, Marracino L, Rapezzi C, Ferrari R. COVID-19 in the heart and the lungs: could we "Notch" the inflammatory storm? *Basic Res Cardiol.* 2020;115(3):31.
33. Liu W, Li H. COVID-19: attacks the 1-beta chain of hemoglobin and captures the porphyrin to inhibit human hem metabolism. *ChemRxiv.* 2019 Dec. Preprint. doi: 10.26434/chemrxiv.11938173.v8.
34. Lippi G, Mattiuzzi C. Hemoglobin value may be decreased in patients with severe coronavirus disease 2019. *Hematol Transfus Cell Ther.* 2020 Apr 2. pii: S2531-1379(20)30029-8. doi: 10.1016/j.htct.2020.03.001.
35. Zhou B, She J, Wang Y, et al. Utility of ferritin, procalcitonin, and c-reactive protein in severe patients. *Research Square.* 2020 Mar 19. Preprint. doi: 10.21203/rs.3.rs-18079/v1.
36. Zheng YY, Ma YT, Zhang JY, Xie X. COVID-19 and the cardiovascular system. *Nat Rev Cardiol.* 2020;17(5):259-60. doi: 10.1038/s41569-020-0360-5.
37. Nataf S. An alteration of the dopamine synthetic pathway is possibly involved in the pathophysiology of COVID-19. *J Med Virol.* 2020 Apr 4. doi: 10.1002/jmv.25826. [Epub ahead of print].
38. Baig AM. Neurological manifestations in COVID-19 caused by SARS-CoV-2. *CNS Neurosci Ther.* 2020;26(5):499-501. doi: 10.1111/cns.13372. Epub 2020 Apr 7.
39. Goh KJ, Choong MC, Cheong EH, et al. Rapid progression to acute respiratory distress syndrome: review of current understanding of critical illness from COVID-19 infection. *Ann Acad Med Singapore.* 2020;49(3):108-118.
40. Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S. Covid-19 does not lead to a "typical" acute respiratory distress syndrome. 2020 Mar 30. doi: 10.1164/rccm.202003-0817LE. [Epub ahead of print].
41. Xu Y, Liu H, Hu K, Wang M. Clinical management of lung cancer patients during the outbreak of 2019 novel coronavirus disease (COVID-19). *Zhongguo Fei Ai Za Zhi.* 2020;23(3):136-141. doi: 10.3779/j.issn.1009-3419.2020.03.02.
42. Recalcati S. Cutaneous manifestations in COVID-19: a first perspective. *J Eur Acad Dermatol Venereol.* 2020 Mar 26. doi: 10.1111/jdv.16387. [Epub ahead of print].
43. Lu D, Sang L, Du S, Li T, Chang Y, Yang XA. Asymptomatic COVID-19 infection in late pregnancy indicated no vertical transmission. *J Med Virol.* 2020 Apr 24. doi: 10.1002/jmv.25927. [Epub ahead of print].
44. Bansal M. Cardiovascular disease and COVID-19. *Diabetes Metab Syndr.* 2020 Mar 25. doi: 10.1016/j.dsx.2020.03.013. [Epub ahead of print].
45. Stebbing J, Phelan A, Griffin I, et al. COVID-19: combining antiviral and anti-inflammatory treatments. *Lancet Infect Dis.* 2020;20(4):400-402. doi: 10.1016/S1473-3099(20)30132-8.
46. Gao J, Tian Z, Yang X. Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. *Biosci Trends.* 2020;14(1):72-73. doi: 10.5582/bst.2020.01047.
47. Touret F, De Lamballerie X. Of Chloroquine and COVID-19. 2020 Mar 5. doi: 10.1016/j.antiviral.2020.104762. [Epub ahead of print].
48. Patri A1, Fabbrocini G. Hydroxychloroquine and ivermectin: a synergistic combination for COVID-19 chemoprophylaxis and/or treatment? *J Am Acad Dermatol.* 2020 Apr 10. pii: S0190-9622(20)30557-0. doi: 10.1016/j.jaad.2020.04.017. [Epub ahead of print].
49. Caly L, Druce JD, Catton MG, Jans DA, et al. The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 *in vitro*. *Antivir Res.* 2020 Apr 3;178:104787. doi: 10.1016/j.antiviral.2020.104787. [Epub ahead of print].
50. Hadadi A, Mortezaadeh M, Kolahehdouzan K, Alavian G. Does recombinant human erythropoietin administration in critically ill COVID-19 patients have miraculous therapeutic effects? *J Med Virol.* 2020 Apr 8. doi: 10.1002/jmv.25839. [Epub ahead of print].
51. Solaimanzadeh I. Acetazolamide, nifedipine and phosphodiesterase inhibitors: rationale for their utilization as adjunctive countermeasures in the treatment of Coronavirus Disease 2019 (COVID-19). *Cureus.* 2020;12(3):e7343. doi: 10.7759/cureus.7343.
52. To KK, Tsang OT, Leung WS, Tam AR, Wu TC, Lung DC, et al. Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: an observational cohort study. *Lancet Infect Dis.* 2020 May;20(5):565-574. doi: 10.1016/S1473-3099(20)30196-1.
53. Luo P, Liu Y, Qiu L, Liu X, Liu D, Li J. Tocilizumab treatment in COVID-19: a single center experience. *J Med Virol.* 2020 Apr 6. doi: 10.1002/jmv.25801. [Epub ahead of print].
54. Bian H, Zheng ZH, Wei D, et al. Meplazumab treats COVID-19 pneumonia: an open-labelled, concurrent controlled add-on clinical trial. *medRxiv.* 2020 Mar 24. Preprint. doi: 10.1101/2020.03.21.20040691.
55. Gautret P, Lagier J, Parola P, Hoang VT, Meddeb L, Mailhe M, Doudier B, Courjon J, Giordanengo V, Vieira VE, Dupont HT, et al. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. *Int J Antimicrob Agents.* 2020 Mar 20. doi: 10.1016/j.ijantimicag.2020.105949. [Epub ahead of print].
56. Zhao J, Yuan Q, Wang H, et al. Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019. *Clin Infect Dis.* 2020 Mar 28. doi: 10.1093/cid/ciaa344. [Epub ahead of print].

57. Bloch EM, Shoham S, Casadevall A, et al. Deployment of convalescent plasma for the prevention and treatment of COVID-19. *J Clin Invest.* 2020 Apr 7. doi: 10.1172/JCI138745. [Epub ahead of print].
58. Xiang F, Wang X1, He X, et al. Antibody Detection and Dynamic Characteristics in Patients with COVID-19. *Clin Infect Dis.* 2020 Apr 19. doi: 10.1093/cid/ciaa461. [Epub ahead of print].
59. Lu X, Xiang Y, Du H, Wing-Kin Wong G, et al. SARS-CoV-2 infection in children – Understanding the immune responses and controlling the pandemic. *Pediatr Allergy Immunol.* 2020 Apr 24. doi: 10.1111/pai.13267. [Epub ahead of print].
60. Hanley B, Lucas SB, Youd E, Swift B, Osborn M. Autopsy in suspected COVID-19 cases. *J Clin Pathol.* 2020 May;73(5):239-242. doi: 10.1136/jclinpath-2020-206522.
61. Fox SE, Akmatbekov A, Harbert JL, Li G, Brown JQ. Pulmonary and cardiac pathology in Covid-19: the first autopsy series from New Orleans. *MedRxiv.* 2020 Apr 10. Preprint. doi: 10.1101/2020.04.06.20050575.
62. Zhang H, Zhou P, Wei Y, et al. Histopathologic changes and SARS-CoV-2 immunostaining in the lung of a patient with COVID-19. *Ann Intern Med.* 2020 Mar 12. doi: 10.7326/M20-0533. [Epub ahead of print].
63. Su H, Yang M, Wan C, et al. Renal histopathological analysis of 26 postmortem findings of patients with COVID-19 in China. *Kidney Int.* 2020 Apr 9. doi: 10.1016/j.kint.2020.04.003. [Epub ahead of print].

Authors' ORCID iDs and academic degrees

Gheorghe Placinta, MD, PhD, Professor of Infectious Diseases – <https://orcid.org/0000-0001-5964-1572>.

Victor Pantea, MD, PhD, Professor of Infectious Diseases – <https://orcid.org/0000-0003-3996-3317>.

Lilia Cojuhari, MD, PhD, Associate Professor of Infectious Diseases – <https://orcid.org/0000-0001-5211-6476>.

Valentin Cebotarescu, MD, PhD, Associate Professor of Infectious Diseases – <https://orcid.org/0000-0003-1089-0038>.

Lidia Placinta, MD, Assistante Professor of Infectious Diseases – <https://orcid.org/0000-0001-9969-867X>.

Dan Croitoru, MD, Undegraduate – <https://orcid.org/0000-0002-8915-0157>.

Authors' contribution

GP conceptualized the project and designed the research; VP revised the manuscript critically. LC interpreted the data; VC performed the laboratory work; LP drafted the manuscript; DC drafted the manuscript and designed the research. All authors revised and approved the final version of the manuscript.

Funding

This study was supported by Research Ethics Committee of *Nicolae Testemitanu* State University of Medicine and Pharmacy. The trial was the authors' initiative. The authors are independent and take responsibility for the integrity of the data and accuracy of the data analysis.

Ethics approval and consent to participate

No approval was required for this study.

Conflict of Interests

No competing interests were disclosed.

