

REVIEW ARTICLES

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A modern approach to interleukin-6

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Abstract

Background: Interleukin-6 is a multifunctional cytokine with well-defined pro- and anti-inflammatory properties. Binding to the receptor complex composed of specific interleukin-6-receptor (IL-6R) and transmembrane glycoprotein gp130, it stimulates various signalling cascades. Intracellular signal transduction involves both STAT-dependent, and STAT-independent mechanisms. IL-6R exists in the soluble and the membrane-bound forms, thus were described the classical signalling and the trans-signalling pathways. The pro-inflammatory response occurs via trans-signalling, while the anti-inflammatory effects are mediated by classical signalling pathway. IL-6 is produced by cardiovascular components. The high levels of IL-6 are identified in the inflammatory diseases.

Conclusions: IL-6 expresses the well-defined pro- and anti-inflammatory properties. The experimental studies have revealed classic signalling pathway with regenerative and anti-inflammatory effects (via the membrane-bound IL-6R), and trans-signalling responsible for the pro-inflammatory response (via the soluble form sIL-6R). The intracellular signal transduction involves the activation of STATs, MAPK, and PI3K cascades. It has been proved that in the cardiovascular pathologies the serum levels of IL-6 correlate with the disease severity and the degree of myocardial damage, being the indicator of heart disease and a predictive factor of adverse outcomes.

Key words: interleukin-6, cytokine, trans-signalling, cardiac ischemia.

Introduction

Cytokines are glycosylated proteins involved in the intercellular communication. The interleukin-6 (IL-6) type of cytokines participates in the inflammation, regulates the target cells differentiation, proliferation, migration and apoptosis [1], and is a key factor in coordinating innate and acquired immune response [2].

Its biological effect is achieved by homo- or heterodimerisation of glycoprotein 130 (gp130) [3]. The transduction of IL-6 type cytokine signal involves the activation of Janus (Jak) kinases, followed by the phosphorylation of transcription factors STAT [4], as well as triggering MAPK and PI3K signalling pathways [5]. The deregulation of both the IL-6-mediated signal transduction mechanism, and the intercellular communication processes generates neoplasms, severe autoimmune and inflammatory diseases [6].

Interleukin-6 (IL-6) is a small glycoprotein that activates the cells via the heterodimeric signalling complex, consisting of the alpha receptor (IL-6R) and the beta subunit for signal transduction, represented by glycoprotein 130 (gp130) common to all cytokines type IL-6 [7].

IL-6-mRNA encodes the 212 amino acid protein, including the 29 amino acid signalling peptide. The secreted protein contains 184 amino acids (21 kDa) [8], of which 107 are well defined in the final structure, while 18 N-end amino acids and 8 amino acids from AB loop have no visible electron density [9]. There have been described the different 21-

28 kDa isoforms, formed due to the various N-linked glycosylation, which determines the stability and half-life of the protein [10]. IL-6 is synthesized and secreted by the most cells, including T cells, fibroblasts, monocytes, and endothelial cells.

The IL-6 conformation consists of 4 long alpha-helices (A, B, C, D) [8], with the typical “up-up-down-down” arrangement, that is common to all IL-6 type cytokines [2]. The attachment to the IL-6R / gp130 receptor complex takes place through three well-differentiated contact sites: *site 1 (binds IL-6R), *site 2 (attaches gp130 between the domains 2 and 3), and *site 3 (contacts the immunoglobulin-like domain 1 of gp130) [2]. The site 1 is formed by the C-end groups of D-helix and the C-end part of the AB loop, and determines the specificity of binding to IL-6R. The site 2, consisting of the middle sequences of the A and C helices, and the site 3, formed by the N-terminal part of the AB loop (3a) and the C-end of the D helix (site 3b), are required for the recruitment of 2 molecules of gp130 [2].

The synthesis of IL-6-mRNA is regulated at both the transcriptional, and the post-transcriptional levels. The key role in IL-6 expression is attributed to nuclear transcription factor kappa B (NF-κB), activated by the bacterial lipopolysaccharides, pro-inflammatory cytokines (TNFα) or viruses [11].

Experimental studies have highlighted fact that a large number of microRNAs (miR) inhibit IL-6 expression, in-

cluding miR-26a [12], miR-142 [13], miR-146a [14], miR-187 [15], miR-200S [16], miR-329 [17].

Very low amounts of IL-6 (about 1-5 µg/ml) there are in the blood of healthy people. The high serum levels of IL-6 are identified in the most of inflammatory and/or autoimmune diseases, reaching the µg/ml values in septic states [18]. IL-6 is the most potent activator of the synthesis of the acute phase proteins in hepatocytes, including the C-reactive protein, and it is an important factor for tumor growth [19]. Kishimoto T. (2010) has suggested that IL-6 is a marker of the continuous inflammation [20].

IL-6 is also produced by cardiovascular components, such as endothelial cells, vascular smooth muscle and ischemic cardiomyocytes [21]. The expression of IL-6 is stimulated by the C-reactive protein via the nuclear factor B (NF-B), and inhibited by the nitric oxide (NO). It was proved the involvement of IL-6 in the cardiac metabolism regulation [22]. Yudkin J. et al. (2000) have shown the role of IL-6 in the pathogenesis and clinical development of atherosclerotic vascular lesions [23].

The high levels of circulating IL-6 are associated with the increased risk of mortality and poor clinical outcome in the patients with unstable angina [24]. Clinically was confirmed that the IL-6 concentrations not only correlate with the severity of the disease, but are also the important predictors of adverse outcomes. It has been shown that the pro-inflammatory cytokines participate in the destabilization and disruption of the atherosclerotic plaque in the coronary arteries, by stimulating the expression of matrix metalloprotease that are responsible for the vascular remodelling and the plaque disorganization [25].

The increased expression of myocardial IL-6 is associated with the progression of heart failure, so IL-6 can be a true indicator of heart damage [26]. The experimental data show evidence that pro-inflammatory cytokines can depress myocardial contractility and are responsible for the cardiovascular disease development and evolution [22]. In the patients with acute coronary syndrome the serum levels of IL-6 correlate with the severity of myocardial lesions [27].

IL-6 is involved in the control of vascular permeability by stimulating the production by the fibroblasts of the vascular endothelial growth factor (VEGF) that acts on the endothelial cells, as well as by enhanced collagen synthesis [28]. There has been identified the association of IL-6 with endothelial cell activation markers, cell adhesion molecules (VCAM1, ICAM1) and von Willebrandt factor [29]. In 2013 Zamani P. et al. (2013) have confirmed that elevated levels of VCAM1 correlate with the increased risk of coronary events in patients with acute coronary syndrome [27].

The published data have proved that IL-6 contributes to the resorption of acute neutrophil infiltration by inducing apoptosis of neutrophils [30]; while the T-cell apoptosis is prevented by activating the STAT3-dependent anti-apoptotic factors (Bcl2, Bclx1) [31].

The *receptor complex* that mediates the biological re-

sponse of IL-6 consists of the transmembrane glycoprotein type I for IL-6 binding, called alpha-IL-6R (CD126 or gp80), and transmembrane protein type I for signal transduction (beta-subunit, CD130 or gp130) [2]. The expression of gp130 is characteristic of all body cell types, whereas the expression of αIL-6R has a limited character and is characteristic of hepatocytes, megakaryocytes, and some leukocyte subpopulations (monocytes, macrophages, B and T cells) [32].

The experimental functional and structural studies suggest that IL-6 can form both the hexameric, composed of 2 molecules of IL-6, IL-6R and gp130 (IL-6₂/IL-6R₂/gp130₂) [33], and the tetrameric (IL-6/IL-6R/gp130₂) signalling complexes [34]. Viswanathan S. et al. (2002) have shown that the low IL-6 concentrations favour the formation of the tetrameric complexes, whereas the high concentrations of IL-6 will lead to the formation of the hexameric complexes [35].

Interleukin-6 receptor (IL-6R) represents the 80 kDa glycosylated membrane protein [36]. The immunoglobulin-like domain of the human IL-6R does not participate in the IL-6 binding, but it is responsible for the receptor stability. The attachment of IL-6 to IL-6R is mediated by the specific sequences located in the domains 2 and 3 of IL-6R [2].

IL-6R exists in both the soluble and the membrane-bound forms, which allow differentiating the classical signalling pathway (via the membrane-bound IL-6R) and the trans-signalling (through soluble (sIL-6R) receptor). In 2012 in his studies, Rose-John S. mentioned that the trans-signalling pathway represents the pro-inflammatory part of the IL-6 biological effect [8], while the classical signalling exhibits the anti-inflammatory and regenerative activity [1]. The experimental studies confirmed that the regenerative, protective and anti-inflammatory effects of IL-6 [37] are mediated through the membrane-bound IL-6R, which is responsible for the differentiation of pro-inflammatory M1 macrophages into anti-inflammatory M2 macrophages [38]. Also via the classical signalling pathway IL-6 is involved in the hepatic regulation of the insulin sensitivity and glucose tolerance [39].

Wilke C. et al. in 2011 highlighted the role of trans-signalling in the immune system adaptation, and the T cells recruitment, activation and apoptosis [40]. The target cells for trans-signalling are the stem cells: hematopoietic [41], nerve [42], smooth muscle [43], and embryonic [44]. The trans-signalling ensures the migration of the lymphocytes in the inflammation area [45]; it induces T cell proliferation and participates in the regulation of the adhesion cells expression in the endothelial cells [46].

IL-6R can be cleaved proteolytically from the cell membrane surface and produce the *soluble form of IL-6R (sIL-6R)* [47]. Taga T. et al. (1989) have demonstrated that the cytosolic region and the transmembrane domain of IL-6 do not participate in the signalling mechanisms, so they have hypothesized that the formation of sIL-6R is a tool for in-

volving the cells that possess only gp130 in the signalling pathways mediated by IL-6 [48].

The soluble sIL-6R receptor is generated by two different mechanisms: * the shedding of the membrane-bound receptor (90-99%) [2, 47]; * the alternative splicing (approximately 1-10%) and the transcription of IL-6R-mRNA omitting the exon 10 that encodes the transmembrane and cytosolic domains [47, 49]. The proteolytic cleavage of IL-6R is catalysed by the ADAM-10 and ADAM-17, Zn²⁺ metalloproteases (shedases) [2, 50], the transmembrane proteins involved in the partial proteolysis of the membrane receptors of the cytokines [51]. The cleavage site for ADAM-17 is located between Gln357 and Asp358 in the region adjacent to cell membrane [52], the complete cleavage of IL-6R from the cell surface occurs within 24 hours [47]. It was supposed that ADAM-10 is responsible for the slow cleavage, whereas ADAM-17 favours the rapid proteolysis of the membrane-bound IL-6R [50]. In addition to the ADAM family of shedases, the IL-6R also can be cleaved by the cathepsin G [53].

The biological activators of ADAM-17 are the pro-inflammatory cytokines (TNF α) [54], reactive C protein, the bacterial toxins (streptolysin O, hemophilin A) and metalloproteases, as well as the apoptotic pathways [55]. In 2011 Scheller J. et al. have mentioned the decisive role of ADAM-17 in the cancer genesis and inflammation [56]. The published data demonstrated that the apoptosis-induced cleavage of IL-6R is caspase-dependent, and PKC/MAPK/ROS independent [2].

According to McFarland-Mancini M. et al. (2010) the main sources of sIL-6R are the hepatocytes and hematopoietic cells, as well as the immune cells (neutrophils, macrophages) [57]. It has been hypothesized that the soluble forms of the receptor are the important regulators of the inflammatory processes [2].

The recent studies have found that the single base pair polymorphism determines the serum level of sIL-6R in humans. The substitution of adenine with cytosine leads to the replacement of Asp358 with Ala358 in the exon 9 of the IL-6 gene [58]. In some studies it has been mentioned the association of Asp358Ala variant with low risk of coronary events [59].

According to Scheller J. et al. (2011) the *glycoprotein 130* (gp130) is a membrane type I glycosylated protein with a molecular weight of 130-150 kDa, consisting of 6 extracellular domains, one transmembrane domain and one cytosolic domain. The N-terminus has the immunoglobulin-like domain (D1), followed by 2 cytokine binding domains (CBD, the domains 2 and 3) and 3 fibronectin (FN-III) domains (the domains 4-6) [2].

In 2003 Boulanger M. et al. demonstrated that the immunoglobulin-like domain of gp130 ensures the interaction with IL-6R and the attachment to the IL-6 site 3 [35]. The cytokine binding domains (CBD2 and CBD3) of gp130 associate with IL-6 site 2. CBD2 located in the N-end region contains 2 pairs of Cys, which form inter-chain disulphide

bonds, while CBD3 contains the Trp-Ser-X-Trp-Ser sequence [60]. The proximal membrane domains of gp130 are involved in the signal transduction to the cytosolic domain [2]. After the ligand binding, the domains 4 and 5 are rearranged at an angle of 80° with the curve formation, thus the whole gp130 ectodomain acquires the structural conformation similar to a wide open «C» [61].

The receptor gp130 subunit has affinity for neither IL-6 nor IL-6R [62]. Therefore, IL-6 can only fix and stimulate the cells that possess IL-6R, while the cells that have only gp130 are completely irresponsive to the IL-6 cytokine signals [63].

The myocardial ischemia/reperfusion induces the expression of gp130, associated with the stimulation of IL-6 and IL-6R expression, thus confirming the role of the IL-6/IL-6R/gp130 system in acute myocardial infarction [12].

Gp130 serves as the β -subunit of the receptor common for the IL-6 cytokine family. Recently have been described several *soluble molecules* (sgp130) with lower molecular mass [64], generated by: (1) shedding, (2) alternative splicing, (3) the addition of an exon consisting of 85 base pairs that alters the codons reading by forming a STOP-codon prior to the transmembrane domain coding sequence, (4) the deletion of an exon with new C-terminal sequence (NIASF) formation, followed by STOP-codon, or (5) the alternative polyadenylation of the intron 10 resulting in new mRNA [65].

Due to the fact that the secreted IL-6 binds in the plasma with sIL-6R, and with sgp130, it has been experimentally proved that the soluble form of gp130 (sgp130) inhibits the trans-signalling pathway [66]. Jostok T. et al. (2001) have confirmed that sgp130 interacts only with sIL-6R in the presence of IL-6. The balance between the soluble and membrane forms of gp130 plays an important role in regulating the biological effects of cytokines. When the amount of IL-6 exceeds the amount of sIL-6R and sgp130, IL-6 will act systemically [62].

The recent publications have demonstrated that the single base pair polymorphism in the IL-6R gene manifested by the replacement of Gln258 with Ala, results in higher serum concentrations of sIL-6R, and is associated with reduced risk of coronary heart disease [67]. Boekholdt S. and Stroes E. (2012) have determined that the cleavage of membrane-bound IL-6R on the surface of hepatocytes, monocytes and macrophages results in the loss of cell sensitivity to IL-6 mediated signals [68]. Scheller J. and Rose-John S. (2012) have assumed that high serum levels of sIL-6R increase the buffer capacity of the sIL-6R/sgp130 complex in the blood, thereby diminishing the systemic effects of IL-6 [69].

There has been shown that the single base pair polymorphism (G148R) in gp130 deregulates the function and stability of the protein, and it is associated with a low risk of acute myocardial infarction [70]. Elevated serum levels of sgp130 were identified in patients with heart failure, being associated with higher mortality rates. In 2007 Ichiki T. et al.

presented the data that in the patients with acute myocardial infarction the serum levels of sgp130 were the highest at admission, and were followed by the decreasing in the post-infarction period [71]. At the same time, the patients with progressive heart failure have higher levels of sgp130 compared to the patients with stable heart failure, so the sgp130 can be used to identify patients at high risk for the heart failure progression [72].

Both the experimental animal studies, and in vitro modelling, have highlighted the beneficial effect of sgp130 in atherosclerosis as result of the trans-signalling pathway inhibition by the specific binding of the IL-6/sIL-6R heterodimer [62, 73]. It has been assumed that the higher levels of sgp130 represent the compensatory response to increased IL-6 signalling in chronic ischemic disease and vascular remodelling [73]. Moreno Velasquez I. et al. (2015) have shown the association of elevated serum levels of sgp130 with a 30% reduction in acute myocardial infarction incidence [74]. There was confirmed the hypothesis that in acute coronary syndromes the sgp130 changes correlate negatively with the severity of the inflammation [75]. The published results have suggested that sgp130 may be used as the prognostic biomarker in cardiac diseases [76].

The classical *signalling pathway* starts with IL-6 attachment to the membrane receptor α IL-6R, the formation of the signalling complex with the gp130 homodimer, the signal transduction and activation of the intracellular signalling pathways [2]. The cells that express only gp130 can be stimulated by the IL-6/sIL-6R complex via the trans-signalling pathway [77].

Cellular response to IL-6/sIL-6R complex differs substantially from the response to IL-6. This can be explained by: * the cell expresses more gp130 than IL-6; ** IL-6 is internalized faster compared to the IL-6/sIL-6R complex [78]. Experimentally has been demonstrated that the IL-6 affinity for IL-6R is 1nM, whereas the affinity of IL-6/sIL-6R complex for gp130 is 100-fold greater [79].

The gp130 cytokine receptor does not possess intrinsic kinase activity. The first step in the intracellular transduction of the IL-6 signal is the activation of Janus (Jak) kinases (Jak1, Jak2 and Tyk2), that are non-covalently constitutive associated with the cytosolic tail of gp130. FERM domain ensures the interaction of Jak-kinases with gp130 [80]. The IL-6 binding to the specific receptor favours the induction of Jak-kinases by the tyrosine sequence phosphorylation, located on the activation loop of C-terminal JH1 domain. The active Jak-kinases lead to phosphorylation of the distal tyrosine residues from the cytosolic part of gp130, which represent the recruitment sites for STAT (signal transducer and activator of transcription) factor. STAT1 or STAT3 monomers attach to the distal phosphotyrosine residue, the tyrosine phosphorylation of STAT occurs, followed by homo- or heterodimerisation, the translocation of STAT dimers in the nucleus, N- and C-terminal acetylation, CBP/p300 interaction, and the attachment to the promotor region of IL-6 in-

ducible gene [81]. Thus, the cytokines binding will induce the reorganization of the preformed receptor complexes and the successive cross-phosphorylation [2].

IL-6 activates the cascades STATs, MAPK, and PI3K [82]. The balance between activation of STAT3 and MAPK is crucial for the controlled proliferation of cells, and ultimately for the body's homeostasis [83]. This balance is regulated by the cytosolic Tyr759 of gp130; the tyrosine phosphorylation will reduce IL-6-dependent STAT3 activation [84], and induce MAPK activation [85]. At the same time, the phosphorylation of the gp130 Tyr759 residue is essential for the membrane IL-6-dependent recruitment of Gab-1 protein, which acts as the bridge between the cascades MAPK, and PI3K, and the activated receptor complex [86]. Chen R. et al. (1999) have concluded that the survival mechanisms are maintained via the activation of PI3K and Akt [87].

Conclusions

Interleukin-6 expresses the well-defined pro- and anti-inflammatory properties. The experimental studies have revealed classic signalling pathway with regenerative and anti-inflammatory effects (via the membrane-bound IL-6R), and trans-signalling responsible for the pro-inflammatory response (via the soluble form sIL-6R). The intracellular signal transduction involves the activation of STATs, MAPK, and PI3K cascades. It has been proved that in the cardiovascular pathologies the serum levels of IL-6 correlate with the disease severity and the degree of myocardial damage, being the indicator of heart disease and a predictive factor of adverse outcomes.

References

- Scheller J, et al. Interleukin-6: From basic biology to selective blockade of pro-inflammatory activities. *Semin Immunol.* 2014 Feb;26(1):2-12.
- Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S. The pro- and anti-inflammatory properties of the cytokine interleukin-6. *Biochem Biophys Acta.* 2011;1813:878-88.
- Heinrich P, Behrmann J, Haan S, Hermanns H, Müller-Newen G, Schaper F. Principles of interleukin (IL)-6-type cytokine signalling and its regulation. *Biochem J.* 2003;374:1-20.
- Akira S. IL-6-regulated transcription factors. *Int J Biochem Cell Biol.* 1997;29:1401-18.
- Eulendorf R, Dittrich A, Khouri C, et al. Interleukin-6 signalling: more than Jaks and STATs. *Eur J Cell Biol.* 2012;91:486-95.
- Dittrich A, et al. Systems biology of IL-6, IL-12 family cytokines. *Cytokine Growth Factor Rev.* 2015;26(5):595-602.
- Wolf J, et al. Interleukin-6 and its receptors: A highly regulated and dynamic system. *Cytokine.* 2014;70:11-20. doi: 10.1016/j.cyto.2014.05.024
- Rose-John S. IL-6 Trans-Signalling via the Soluble IL-6 Receptor: Importance for the pro-inflammatory activities of IL-6. *Int J Biol Sci.* 2012;8(9):1237-47.
- Somers W, Stahl M, Seehra JS. A crystal structure of interleukin 6: implications for a novel mode of receptor dimerization and signalling. *EMBO J.* 1997;16(5):989-97.
- Schiel X, Rose-John S, Duffhues G, Schooltink H, Gross V, Heinrich PC. Microheterogeneity of human interleukin 6 synthesized by transfected NIH/3T3 cells: comparison with human monocytes, fibroblasts and endothelial cells. *Eur J Immunol.* 1990;20(4):883-7.

11. Mansell A, Jenkins BJ. Dangerous liaisons between interleukin-6 cytokine and toll-like receptor families: a potent combination in inflammation and cancer. *Cytokine Growth Factor Rev.* 2013;24(3):249-56.
12. Yang X, Liang L, Zhang X-FF, Jia H-LL, Qin Y, Zhu X-CC, et al. MicroRNA-26a suppresses tumor growth and metastasis of human hepatocellular carcinoma by targeting Interleukin-6-Stat3 pathway. *Hepatology.* 2013;58(1):158-70.
13. Sun Y, Sun J, Tomomi T, Nieves E, Mathewson N, Tamaki H, et al. PU.1-dependent transcriptional regulation of miR-142 contributes to its hematopoietic cell-specific expression and modulation of IL-6. *J Immunol.* 2013;190(8):4005-13.
14. He Y, Sun X, Huang C, Long XR, Lin X, Zhang L, et al. MiR-146a regulates IL-6 production in lipopolysaccharide-induced RAW264 7 macrophage cells by inhibiting Notch1. *Inflammation.* 2014;37(1):71-82.
15. Rossato M, Curtale G, Tamassia N, Castellucci M, Mori L, Gasperini S, et al. IL-10-induced microRNA-187 negatively regulates TNF- α , IL-6, and IL-12p40 production in TLR4-stimulated monocytes. *Proc Natl Acad Sci USA.* 2012;109(45):E3101-10.
16. Dou L, Zhao T, Wang L, Huang X, Jiao J, Gao D, et al. MiR-200s contribute to Interleukin-6 (IL-6)-induced insulin resistance in hepatocytes. *J Biol Chem.* 2013; 288(31):22596-606.
17. Garg M, Potter JA, Abrahams VM. Identification of microRNAs that regulate TLR2-mediated trophoblast apoptosis and inhibition of IL-6 mRNA. *PLoS ONE.* 2013;8(10):e77249.
18. Waage A, Brandtzaeg P, Halstensen A, Kierulf P, Espevik T. The complex pattern of cytokines in serum from patients with meningococcal septic shock. *J Exp Med.* 1989;169(1):333-8.
19. Taniguchi K, Karin M. IL-6 and related cytokines as the critical lynchpins between inflammation and cancer. *Semin. Immunol.* 2014;26(1):54-74.
20. Kishimoto T. IL-6: from its discovery to clinical applications. *Int Immunol.* 2010;22(5):347-52.
21. Gwechenberger M, Mendoza LH, Youker KA, et al. Cardiac myocytes produce interleukin-6 in culture and in viable border zone of reperfused infarctions. *Circulation.* 1999;99(4):546-51.
22. Kanda T, Takahashi T. Interleukin-6 and Cardiovascular Diseases. *Jpn Heart J.* 2004;45(2):183-93.
23. Yudkin JS, Kumari M, Humphries SE, et al. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis.* 2000;148(2):209-14.
24. Biasucci LM, Liuzzo G, Fantuzzi G, et al. Increasing levels of interleukin (IL)-1Ra and IL-6 during the first 2 days of hospitalization in unstable angina are associated with increased risk of in-hospital coronary events. *Circulation.* 1999;99(16):2079-84.
25. Rajavashisth TB, Xu XP, Jovinge S, et al. Membrane type 1 matrix metalloproteinase expression in human atherosclerotic plaques: evidence for activation by proinflammatory mediators. *Circulation.* 1999;99(24):3103-9.
26. Plenz G, Song ZF, Reichenberg S, et al. Left-ventricular expression of interleukin-6 messenger-RNA higher in idiopathic dilated than in ischemic cardiomyopathy. *Thorac Cardiovasc Surg.* 1998;46(4):213-6.
27. Zamani P, Schwartz GG, Olsson AG, et al.; Myocardial Ischemia Reduction with Aggressive Cholesterol Lowering (MIRACL) Study Investigators. Inflammatory biomarkers, death, and recurrent nonfatal coronary events after an acute coronary syndrome in the MIRACL study. *J Am Heart Assoc.* 2013;2(1):e003103.
28. Nakahara H, et al. Anti-interleukin-6 receptor antibody therapy reduces vascular endothelial growth factor production in rheumatoid arthritis. *Arthritis Rheum.* 2003;46(8):1521-9.
29. Mulvihill NT, Foley JB, Murphy RT, et al. Risk stratification in unstable angina and non-Q wave myocardial infarction using soluble cell adhesion molecules. *Heart.* 2001;85(6):623-7.
30. Kaplanski G, Marin V, Montero-Julian F, Mantovani A, Farnarier C. IL-6: a regulator of the transition from neutrophil to monocyte recruitment during inflammation. *Trends Immunol.* 2003;24(1):25-9.
31. Jones SA. Directing transition from innate to acquired immunity: defining a role for IL-6. *J Immunol.* 2005;175(6):3463-8.
32. Oberg HH, Wesch D, Grussel S, Rose-John S, Kabelitz D. Differential expression of CD126 and CD130 mediates different STAT-3 phosphorylation in CD4+CD25- and CD25 high regulatory T cells. *Int Immunol.* 2006;18(4):555-63.
33. Boulanger MJ, Chow DC, Brevnova EE, Garcia KC. Hexameric structure and assembly of the interleukin-6/IL-6 alpha receptor/gp130 complex. *Science.* 2003;300:2101-4.
34. Grötzinger J, Kernebeck T, Kallen K-J, Rose-John S. IL-6 type cytokine receptor complexes: hexamer or tetramer or both? *Biol Chem.* 1999;380(7-8):803-13.
35. Viswanathan S, Benatar T, Rose-John S, Lauffenburger DA, Zandstra PW. Ligand/ receptor signalling threshold (LIST) model accounts for gp130-mediated embryonic stem cell self-renewal responses to LIF and HIL-6. *Stem Cells.* 2002;20(2):119-38.
36. Rincon M. Interleukin-6: from an inflammatory marker to a target for inflammatory diseases. *Trends Immunol.* 2012;33(11):571-7. doi: 10.1016/j.it.2012.07.003.
37. Luig M, Kluger MA, Goerke B, Meyer M, Nosko A, Yan I, et al. Inflammation-induced IL-6 functions as a natural brake on macrophages and limits GN. *J Am Soc Nephrol.* 2015;26(7):1597-607.
38. Mauer J, Chaurasia B, Goldau J, Vogt MC, Ruud J, Nguyen KD, et al. Signalling by IL-6 promotes alternative activation of macrophages to limit endotoxemia and obesity-associated resistance to insulin. *Nat Immunol.* 2014;15(5):423-30.
39. Febbraio MA, Rose-John S, Pedersen BK. Does interleukin-6 receptor blockade the Holy Grail for inflammatory diseases? *Clin Pharmacol Ther.* 2010;87(4):396-8.
40. Wilke CM, Bishop K, Fox D, Zou W. Deciphering the role of Th17 cells in human disease. *Trends Immunol.* 2011;32(12):603-11.
41. Audet J, Miller CL, Rose-John S, Piret JM, Eaves CJ. Distinct role of gp130 activation in promoting self-renewal divisions by mitogenically stimulated murine hematopoietic cells. *Proc Natl Acad Sci USA.* 2001;98(4):1757-62.
42. März P, Heese K, Dimitriadis-Schmutz B, Rose-John S, Otten U. Role of interleukin-6 and soluble IL-6 receptor in region specific induction of astrocytic differentiation and neurotrophin expression. *Glia.* 1999;26(3):191-200.
43. Klouche M, Bhakdi S, Hemmes M, Rose-John S. Novel Path of activation of primary human smooth muscle cells: upregulation of gp130 creates an autocrine activation loop by IL-6 and its soluble receptor. *J Immunol.* 1999;163(8):4583-9.
44. Humphrey RK, Beattie GM, Lopez AD, Bucay N King CC, Firpo M, et al. Maintenance of pluripotency in human embryonic stem cells is Stat3 independent. *Stem cells.* 2004;22(4):522-30.
45. Rabe B, Chalaris A, May, Waetzig GH, Seeger D, Williams AS, Jones SA, Rose-John S, Scheller J. Transgenic blockade of interleukin 6 trans-signalling abrogates inflammation. *Blood.* 2008;111(3):1021-8.
46. Chen Q, Fisher DT, Clancy KA, Gauguet JM, Wang WC, Unger E, Rose-John S, von Andrian UH, Baumann H, Evans S.S. Fever-range thermal stress promotes lymphocyte trafficking across high endothelial venules via an interleukin 6 trans-signalling mechanism. *Nat Immunol.* 2006;7(12):1299-1308.
47. Müllberg J, Schooltink H, Stoyan T, Gunther M, Graeve L, Buse G, et al. The soluble interleukin-6 receptor is generated by shedding. *Eur J Immunol.* 1993;23(2):473-80.
48. Taga T, Hibi M, Hirata Y, Yamasaki K, Yasukawa K, Matsuda T, et al. Interleukin-6 triggers the association of its receptor with a possible signal transducer, gp130. *Cell.* 1989;58(3):573-81.
49. Lust JA, Donovan KA, Kline MP, Greipp PR, Kyle RA, Maihle NJ. Isolation of a mRNA encoding a soluble form of the human interleukin-6 receptor. *Cytokine.* 1992;4(2):96-100.
50. Matthews V, Schuster B, Schutze S, Bussmeyer I, Ludwig A, Hundhausen C, et al. Cellular cholesterol depletion triggers shedding of the human Interleukin-6 receptor by ADAM10 and ADAM17 (TACE). *J Biol Chem.* 2003;278(40):38829-39.
51. Black RA. Tumor necrosis factor-alpha converting enzyme. *Int J Biochem Cell Biol.* 2002;34(1):1-5.
52. Müllberg J, Oberthür W, Lottspeich F, Mehl E, Dittrich E, Graeve L, et al. The soluble human IL-6 receptor. Mutational characterization of the proteolytic cleavage site. *J Immunol.* 1994;152(10):4958-68.
53. Bank U, Reinhold D, Schneemilch C, Kunz D, Synowitz H, Ansgorge S. Selective proteolytic cleavage of IL-2 receptor and IL-6 receptor

- ligand binding chains by neutrophil-derived serine proteases at foci of inflammation. *J Interferon Cytokine Res.* 1999;19(11):1277–87.
54. Franchimont N, Lambert C, Huynen P, Ribbens C, Relic B, Chariot A, Bours V, Piette J, Merville MP, Malaise M. Interleukin-6 receptor shedding is enhanced by interleukin-1beta and tumor necrosis factor alpha and is partially mediated by tumor necrosis factor alpha-converting enzyme in osteoblast-like cells. *Arthritis Rheum.* 2005;52(1):84–93.
 55. Chalaris A, Rabe B, Paliga K, Lange H, Laskay T, Fielding CA, et al. Apoptosis is a natural stimulus of IL6R shedding and contributes to the pro-inflammatory trans-signalling function of neutrophils. *Blood.* 2007;110(6):1748–55.
 56. Scheller J, Chalaris A, Garbers C, Rose-John S. ADAM17: a molecular switch of inflammatory and regenerative responses? *Trends Immunol.* 2011;32(8):380–7.
 57. McFarland-Mancini MM, Funk HM, Paluch AM, Zhou M, Giridhar PV, Mercer CA, Kozma SC, Drew AF. Differences in wound healing in mice with deficiency of IL-6 versus IL-6 receptor. *J Immunol.* 2010;184(12):7219–28.
 58. Galicia J, Tai H, Komatsu Y, Shimada Y, Akazawa K, Yoshie H. Polymorphisms in the IL-6 receptor (IL-6R) gene: strong evidence that serum levels of soluble IL-6R are genetically influenced. *Genes Immun.* 2004;5(6):513–6.
 59. Collaboration IRGCERF, Sarwar N, Butterworth AS, Freitag DF, Gregson J, Willeit P, et al. Interleukin-6 receptor pathways in coronary heart disease: a collaborative meta-analysis of 82 studies. *Lancet.* 2012;379:1205–13.
 60. Bazan JF. Structural design and molecular evolution of a cytokine receptor superfamily. *Proc Natl Acad Sci USA.* 1990;87(18):6934–8.
 61. Lorenzen I, Shang W, Perbandt M, Petoukhov MV, Svergun D, Waetzig GH, Rose-John S, Hilgenfeld R, Grötzinger J. The structure of the unliganded extracellular domain of the interleukin-6 signal transducer gp130 in solution. *Eur J Cell Biol.* 2011;90(6-7):515–20.
 62. Jostock T, Müllberg J, Özbek S, Atreya R, Blinn G, Voltz N, et al. Soluble gp130 is the natural inhibitor of soluble IL-6R trans-signalling responses. *Eur J Biochem.* 2001;268(1):160–7.
 63. Fischer M, Goldschmitt J, Peschel C, Kallen KJ, Brakenhoff J, Wollmer A, et al. A designer cytokine with high activity on human hematopoietic progenitor cells. *Nat Biotech.* 1997;15(2):142–5.
 64. Tanaka M, Kishimura M, Ozaki S, Osakada F, Hashimoto H, Okubo M, Murakami M, Nakao K. Cloning of novel soluble gp130 and detection of its neutralizing autoantibodies in rheumatoid arthritis. *J Clin Invest.* 2000;106(1):137–44.
 65. Diamant M, Rieneck K, Mechti N, Zhang XG, Svenson M, Bendtzen K, et al. Cloning and expression of an alternatively spliced mRNA encoding a soluble form of the human Interleukin-6 signal transducer gp130. *FEBS Lett.* 1997;412(2):379–84.
 66. Richards PJ, Nowell MA, Horiuchi S, McLoughlin RM, Fielding CA, Grau S, et al. Functional characterization of a soluble gp130 isoform and its therapeutic capacity in an experimental model of inflammatory arthritis. *Arthritis Rheum.* 2006;54(5):1662–72.
 67. Hingorani AD, Casas JP. The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomization analysis. *Lancet.* 2012;379(9822):1214–24.
 68. Boekholdt SM, Stroes ES. The interleukin-6 pathway and atherosclerosis. *Lancet.* 2012;379(9822):1176–8.
 69. Scheller J, Rose-John S. The interleukin 6 pathway and atherosclerosis. *Lancet.* 2012;380(9839):338.
 70. Benrick A, Jirholt P, Wernstedt I, Gustafsson M, Scheller J, Eriksson AL, et al. A non-conservative polymorphism in the IL-6 signal transducer (IL6ST)/gp130 is associated with myocardial infarction in a hypertensive population. *Regul Pept.* 2008;146(1-3):189–96.
 71. Ichiki T, Jougasaki M, Setoguchi M, Shimokawahara H, Nakashima H, Matsuoka T, et al. Plasma levels of soluble glycoprotein 130 in acute myocardial infarction. *J Cardiol.* 2007;50(2):101–9.
 72. Gwechenberger M, Pacher R, Berger R, Zorn G, Moser P, Stanek B, et al. Comparison of soluble glycoprotein 130 and cardiac natriuretic peptides as long-term predictors of heart failure progression. *J Heart Lung Transplant.* 2005;24(12):2190–5.
 73. Morieri ML, Passaro A, Zuliani G. Interleukin-6 (Trans-Signalling) and ischemic vascular disease: the important role of soluble gp130. *Mediators Inflamm.* 2017;2017:1396398.
 74. Moreno Velasquez I, Golabkesh Z, K'allberg H, Leander K, de Faire U, Gigante B. Circulating levels of interleukin 6 soluble receptor and its natural antagonist, sgp130, and the risk of myocardial infarction. *Atherosclerosis.* 2015;240(2):477–81.
 75. Ritschel VN, Seljeflot I, Arnesen H, et al. Circulating levels of IL-6 receptor and gp130 and long-term clinical outcomes in ST-elevation myocardial infarction. *J Am Heart Assoc.* 2016;5(6). pii: e003014.
 76. Askevold ET, Nymo S, Ueland T, Gravning J, Wergeland R, Kjekshus J, et al. Soluble glycoprotein 130 predicts fatal outcomes in chronic heart failure: analysis from the Controlled Rosuvastatin Multinational Trial in Heart Failure (CORONA). *Circ Heart Fail.* 2013;6(1):91–8.
 77. Rose-John S, Waetzig GH, Scheller J, Grotzinger J, Seegert D. The IL-6/sIL-6R complex as a novel target for therapeutic approaches. *Expert Opin Ther Targets.* 2007;11(5):613–24.
 78. Peters M, Blinn G, Solem F, Fischer M, Meyer zum Büschenfelde KH, Rose-John S. In vivo and in vitro activity of the gp130 stimulating designer cytokine Hyper-IL-6. *J Immunol.* 1998;161(7):3575–81.
 79. Rose-John S, Schooltink H, Lenz D, Hipp E, Dufhues G, Schmitz H, et al. Studies on the structure and regulation of the human hepatic interleukin-6 receptor. *Eur J Biochem.* 1990;190(1):79–83.
 80. Haan C, Heinrich PC, Behrmann I. Structural requirements of the interleukin-6 signal transducer gp130 for its interaction with Janus kinase 1: the receptor is crucial for kinase activation. *Biochem J.* 2002;361:105–11.
 81. Ray S, Boldogh I, Brasier AR. STAT3 NH2-terminal acetylation is activated by the hepatic acute-phase response and required for IL-6 induction of angiotensinogen. *Gastroenterology.* 2005;129(5):1616–32.
 82. Eulendorf R, Dittrich A, Khouri C, Müller PJ, Müller B, Wolf A, et al. Interleukin-6 signalling: more than Jaks and STATs. *Eur J Cell Biol.* 2012;91(6-7):486–95.
 83. Fukada T, Ohtani T, Yoshida Y, Shirogane T, Nishida K, Nakajima K, et al. STAT3 orchestrates contradictory signals in cytokine-induced G1 to S cell-cycle transition. *EMBO J.* 1998;17(22):6670–7.
 84. Schaper F, Gendo C, Eck M, Schmitz J, Grimm C, Anhof D, et al. Activation of the protein tyrosine phosphatase SHP2 via the interleukin-6 signal transducing receptor protein gp130 requires JAK1 and limits acute-phase protein expression. *Biochem J.* 1998;335:557–65.
 85. Lai CF, Ripperger J, Wang Y, Kim H, Hawley RB, Baumann H. The STAT3-independent signalling pathway by glycoprotein 130 in hepatic cells. *J Biol Chem.* 1999;274(12):7793–802.
 86. Eulendorf R, Schaper F. A new mechanism for the regulation of Gab1 recruitment to the plasma membrane. *J Cell Sci.* 2009;122:55–64.
 87. Chen RH, et al. Interleukin-6 inhibits transforming growth factor-beta-induced apoptosis through the phosphatidylinositol 3-kinase/Akt and signal transducers and activators of transcription 3 pathways. *J Biol Chem.* 1999;274(33):23013–9.